# MANTER: Journal of Parasite Biodiversity (ISSN 2470-8224) Occasional Papers, Number 3, August 31, 2016. doi:10.13014/K27P8W9Z Copyright © 2016 Mauldin, Doty, Nakazawa, Emerson, and Carroll.



This paper was part of a symposium on mammal parasite biodiversity, "CLM20 — Zoonosis y mamíferos Neotropicales" [Zoonoses and Neotropical Mammals], presented at III Congreso Latinoamericano de Mastozoología, Bogotá D.C., Colombia, 1 al 5 de diciembre del 2015.

# The Importance of Mammalogy, Infectious Disease Research, and Biosafety in the Field

Matthew R. Mauldin,<sup>1,2</sup> Jeffrey B. Doty,<sup>1</sup> Yoshinori Nakazawa,<sup>1</sup> Ginny L. Emerson,<sup>1</sup> and Darin S. Carroll<sup>3</sup>

- 1 Poxvirus and Rabies Branch, Centers for Disease Control and Prevention, 1600 Clifton Road NE, Atlanta, Georgia, USA
- 2 Oak Ridge Institute for Science and Education (ORISE), CDC Fellowship Program, Oak Ridge, Tennessee, USA
- 3 Environmental, Safety, and Health Compliance Office, Centers for Disease Control and Prevention, 1600 Clifton Road NE, Atlanta, Georgia, USA

Corresponding author - M. R. Mauldin, email mmauldin@cdc.gov

#### **Abstract**

Large amounts of data and multitudes of publications have been independently generated by researchers in mammalogy and infectious diseases. The frequent confluence of these fields in epidemiological research as well as the facility of the data generated to be used in applied methods (e.g., conservation, public outreach, public health interventions) suggests that the intersection of these fields is important not only to their committed scientists but also to other areas of investigation, including public health. Given the increased frequency with which researchers in these fields interact with potentially infected humans, animals, and tissues, their occupations present a higher risk of exposure to a variety of pathogens than those in other fields of biology or among most jobs of the general public. However, a variety of methods are available for minimizing this risk, including increasing awareness of potential risks, using medical prophylaxes (when available), properly employing personal protective equipment, and using adequate disinfectants. Although instances of serious illness from zoonotic diseases among field researchers may be uncommon, they do occur; the purpose of this document is to increase awareness of risks that researchers—principal investigators and students alike—face and highlight steps and resources that can mitigate those risks.

Keywords: mammalogy, personal protective equipment, precautions, risks, safety, zoonoses

### Introduction

Mammalogy is a broad field of study that includes such diverse disciplines as studying morphological or physiological variability (Gardner et al., 2014), understanding animal behavior (Davis et al., 1999), examining levels and patterns of genetic variability (Olayemi et al., 2012; Bradley and Mauldin, 2016), and monitoring overall trends in mammalian biodiversity (Willig et al., 2003). These disciplines provide the scientific community with a better understanding of the ecology and evolution of mammals, further improving our understanding of biology for all taxa. Previous studies have also shown that basic research often can have economic importance (Salter and Martin, 2001).

To mammalogists, the increased knowledge of the natural world as well as potential economic benefits of research adequately illustrate the importance of mammalogy; however, the increased understanding of mammalian evolution and biodiversity can also influence various other fields of study, including public education (Gore et al., 2006) and wildlife conservation (Young, 1994).

Multiple studies have illustrated the high diversity of mammalian species present in the lower latitudes (Kaufman, 1995; Willig et al., 2003; Ceballos and Ehrlich, 2006). A recent publication ranked the top 19 countries with the greatest known diversity of mammalian species, and each one contains at least some landmass in the tropics (Ceballos, 2014). Given these data, it should not be surprising that the

tropics are a hotbed of biological research. Additionally, this is an area of constant discovery, and the rate at which new mammal species are being identified has increased since the 1960s; furthermore, a greater number of recently identified mammalian species have been reported from the tropics than temperate regions (Reeder et al., 2006). In the same period of time, many areas of high biodiversity and endemism (in the tropics and elsewhere) have lost increasingly large portions of primary habitat because of anthropogenic activity, endangering known endemic species as well as those still unknown to science (Ceballos and Ehrlich, 2006; Myers et al., 2000). This decline highlights the need and importance for continued research of mammalian biodiversity, public education, and conservation in the tropics as well as in other parts of the world.

Researchers working with emerging infectious disease (EID) have reported similar geographical (the lower latitudes) and temporal trends (increased rate of discovery) (Jones et al., 2008). Studies have indicated that the frequency of EID events has increased since the 1940s, suggesting the threat of EIDs to global health is increasing (Jones et al., 2008; Brooks et al., 2014). Furthermore, it has been estimated that approximately 60% of EIDs are zoonotic in nature (Jones et al., 2008), and that zoonotic diseases, both emerging and established, cause an estimated 2.5 billion human illnesses and 2.7 million deaths each year (Grace et al., 2012). Given the global distribution of zoonoses along with their substantial human health implications, zoonotic disease research is an extremely active field with a wide variety of disciplines. Some research efforts have utilized algorithms to model the ecological niche of a disease (Peterson 2006) or the spread of infection (Meyers, 2007). Researchers often use genetic data to examine phylogeographic and evolutionary patterns of pathogens (Nakazawa et al., 2015); others conduct vaccine efficacy testing (Agnandji et al., 2016; Keckler et al., 2011) or examine cellular mechanisms used by antivirals (Hemmi et al., 2002).

Serological surveys have helped identify both potential reservoir/host species of zoonoses and prevalence of infection in sylvatic or urban systems (Field et al., 2001; Pitts et al., 2013; Nolen et al., 2015). When a potential mammalian reservoir is identified based on serological surveys or other investigations, mammalogists may know a great deal about the host, but a paucity of information regarding its taxonomy and/or phylogeographic patterns could be just as likely. One case of the former was the identification of *Peromyscus maniculatus* (deer mouse) as the primary reservoir of the hantavirus *Sin Nombre virus* (SNV) (Nichol et al., 1993; Childs et al., 1994). The genus *Peromyscus* is one of the most actively studied genera of North America, and this research provided information regarding the geographic

distribution and ecology of the SNV reservoir, which was used to better determine additional regions and habitats that might harbor both the reservoir and virus. An example of the latter is the identification of Cricetomys spp. as a potential reservoir for Monkeypox virus (MPXV) through serological surveys as well as its involvement in the 2003 outbreak in the United States (Hutin et al., 2001; Hutson et al., 2007; Reynolds et al., 2010). Although the geographic distribution of the genus as a whole is fairly well understood, there is uncertainty in the number of recognized species. Four species are recognized by Musser and Carleton (2005), yet the most recent publication of the IUCN red list (van der Straeten et al., 2008) recognized only two species. All studies available for the listed citations relied upon morphological and ecological data. The most extensive molecular examination of the genus published to date recognized three additional lineages that likely represent previously unrecognized species (Olayemi et al., 2012), and ongoing research has identified an additional two lineages, suggesting the number of species is as high as eight (Mauldin et al., unpublished data). Those genetic analyses indicate the presence of an eastern and western clade of Cricetomys, similar to the geographic pattern seen in MPXV (West African and Congo Basin clades; Likos et al., 2005; Nakazawa et al., 2015), although previous understanding suggested the distributions of Cricetomys gambianus and Cricetomys emini stretched from west to east Africa. Identification of geographic barriers to gene flow of mammalian reservoirs has the potential to provide insights into geographic structure of the virus as well. This is especially interesting within the MPXV system, given the difference in virulence between the West African and Congo Basin clades of MPXV.

Through both mammalian and epidemiological studies, vast amounts of data have been generated and are frequently used to guide public health interventions to save lives and improve the quality of life for people around the globe. After the discovery of SNV in the United States, research conducted by mammalogists and epidemiologists led to a better understanding of the genetic and geographic variability of SNV and the host relationships of the species of New World hantaviruses (Nichol et al., 1993; Childs et al., 1994; Fulhorst et al., 2007; Pitts et al., 2013; Montoya-Ruiz et al., 2014). These studies prompted discussion and changes in safety regulations that affected many researchers (Fulhorst et al., 2007; Mills et al., 1995a; Mills et al., 1995b; Kelt et al., 2007). Similar research has illuminated biogeographic patterns of pathogenic South American arenaviruses (Griffiths et al., 1992; Fulhorst et al., 1997; Delgado et al., 2008; Irwin et al., 2012).

Historically, understanding routes of infection and how to minimize the risk of human exposure through large-scale vaccination campaigns led to the elimination of canine rabies from the United States. This, along with the implementation of oral rabies vaccine for geographic containment of host-associated rabies strains and the development of post-exposure prophylaxis (PEP) has decreased the number of annual human rabies deaths in the United States from approximately 100 in the early 1900s to about 2 today (Finnegan et al., 2002). Globally, an estimated 15 million people receive rabies PEP annually, which is thought to save hundreds of thousands of lives each year (WHO, 2005). These life-saving applications would not have been possible without epidemiological research to understand the natural history and modes of transmission for hantaviruses, or the host-specific and geographically distinct patterns of Rabies virus variants throughout North America. These data were then utilized for public outreach and the design of oral rabies vaccine campaigns.

Similar success stories are many, regarding both zoonotic and human-specific pathogens, and countless lives have been saved throughout the world. Unfortunately, there are still many diseases, specifically in the tropics (commonly referred to as "neglected tropical diseases"), which affect billions of people, primarily in developing countries (Feasey et al., 2009). Jones et al. (2008) determined that the risk of wildlife, zoonotic, and vector-borne EIDs originating at the lower latitudes is substantial and suggested better allocation of global resources to improve EID research in the tropics. As zoonoses are capable of infecting both humans and wildlife, the finding that mammalian biodiversity is a strong predictor of disease co-occurrence (Hoberg and Brooks, 2015; Murray et al., 2015) is not surprising. This correlation and the trend of increased mammalian biodiversity in the lower latitudes further supports the need for research in both mammalian diversity and zoonotic diseases in the tropics.

## Few Things Worth Doing Are without Risk

Clearly, as discussed earlier, there are many benefits to both mammalogical research and zoonotic disease research, including the generation of valuable information regarding evolution, speciation, and the ecology of their respective taxa. Additionally, these data have been used in applied fields such as conservation and mitigating risk of zoonotic diseases, among countless other implementations. However, researchers (e.g., wildlife biologists, pathologists, epidemiologists, and public health officials) and others working with potentially infected people, animals, and materials, are at a higher risk of contracting zoonotic diseases than is the general population (Zeitz et al., 1995; Mann et al., 1984; Baker and Gray, 2009). Mammalogists may not intention-

ally seek out infected animals, but many work in potential hotspots for EIDs or with mammals known to harbor various pathogens, placing them at higher risk of exposure to certain zoonoses.

Researchers have used serosurvey results and questionnaire responses from a variety of professional meetings (e.g., American Society of Mammalogists, Wildlife Disease Association, Southwestern Association of Naturalists) to argue both for (Fulhorst et al., 2007) and against (Kelt et al., 2007) the need for additional personal protective equipment (PPE) in the field determined by assessing the risk of occupational exposure to specific pathogens. Although the proportion of researchers exhibiting evidence of previous exposure to disease-causing pathogens may be low, there is a very real risk in working with pathogens or pathogen-associated organisms (mammals and vectors). There are multiple instances of biologists working in the field (either handling animals directly, entering areas inhabited by mammals, or simply exploring nature) becoming ill and even dying as a result of exposure to zoonotic diseases. Although some researchers in the following examples were investigating pathogens, others were not.

In 2004 a wildlife sciences graduate student working in West Virginia was infected with the Monongahela virus (a species of hantavirus), developed symptoms of HPS, and died shortly after hospitalization (Sinclaire et al., 2007). Lack of PPE when handling rodents and a failure to wash hands before eating were referenced as two potential exposures to the virus. The same study reported a retrospectively diagnosed 1981 case of HPS in a wildlife biologist who worked in one of the same counties (Sinclaire et al., 2007). Also in 2004, a field technician became ill with HPS in California, although it was undetermined whether the exposure occurred from direct handling of small mammals or from sleeping in a bunkhouse with rodents that tested positive for antibodies reactive to SNV (Kelt et al., 2007). One year later, two field workers became ill and were hospitalized in Boulder, Colorado, after collecting rodents for ecological studies at separate field localities (Torres-Pérez et al., 2010). A wildlife biologist died from the plague in 2007 in Arizona after conducting a necropsy on a mountain lion without the use of proper PPE (Wong et al., 2009). In 2008 two American scientists contracted Zika virus while working in Senegal, and person-to-person transmission was proposed when the wife of one scientist also became ill (Foy et al., 2011). In 2011, 13 biology students developed an acute respiratory illness (some were hospitalized). All are suspected to have acquired pulmonary histoplasmosis from entering a hollow, bat-infested tree in Uganda during a class field trip. Serological data confirmed diagnosis for five of the students (Cottle et al., 2013).

These are examples of infection from recognized pathogens known to occur in those areas; however, with the increased rate of EIDs, more and more diseases are being discovered (Brooks et al., 2014). One such instance was the discovery of Sosuga virus when a young biologist became infected with a novel paramyxovirus after fieldwork in South Sudan and Uganda. This biologist was hospitalized for two weeks with severe acute febrile disease and had lingering symptoms for months (Albariño et al., 2014; Amman et al., 2015). The exact time and location of exposure is unknown; however, it appears inconsistent adherence to use of proper PPE may have occurred. Vector-borne diseases are also a health concern, as one wildlife biologist is reported to have contracted Lyme disease three times (Moyer 2015). Collectively, these incidents occurred over wide latitudinal and elevational gradients, indicating the need for biosafety and risk mitigation in the field regardless of where fieldwork is conducted.

The point of discussing examples of field biologists becoming ill from a variety of pathogens is not an attempt to scare and manipulate researchers into unnecessarily increasing their use of PPE at the cost of comfort and mobility. The authors acknowledge that these instances are rare considering the number of researchers working in the field around the world, and many mammalogists as well as wildlife biologists have admitted to rarely (if ever) using gloves, respiratory protection, or splash guards (Fulhorst et al., 2007; Kelt et al., 2007; Bosch et al., 2013) and have no detected history of zoonotic disease infection. Rather, the reason is to remind principal investigators, as well as those entering the realm of field biology, that there are inherent risks associated with handling small mammals and to initiate a conversation regarding the most appropriate and efficient ways to mitigate those risks. It is the hope of the authors that this discussion will help researchers make informed decisions regarding the health of themselves, their colleagues, and students.

# PPE and Other Ways to Mitigate Risk of Exposure

It has been suggested that "the most important prophylactic measure for personnel who are trapping, handling, bleeding, or dissecting rodents is to be mindful of potential routes of infection and carefully avoid conditions which may lead to transmission" (Mills et al., 1995a; CDC, 1993). This practice begins with being aware of dangers and informing all participants of potential risks and methods that can minimize those risks (Mills et al., 1995a). Risk mitigation should commence well before actual fieldwork and include educating all participants of potential zoonoses or infec-

tious diseases known to occur in the area (see CDC Yellow Book; CDC, 2016), symptoms of those diseases, and ways to avoid becoming ill (including suggested medical prophylaxes, if available) as well as proper training regarding the use of PPE, animal handling, and necropsy techniques. Discussion should also include that active handling is not the only method of potential exposure and that safety should be considered first in all field activities, including the avoidance of housing that shows evidence of occupation by rodents (Kelt et al., 2010) and not entering enclosed, poorly ventilated areas (i.e., abandoned buildings, caves, attics, hollow trees, etc.) without appropriate respiratory protection because even short exposure times in enclosed areas can result in infection with some zoonotic diseases (Tsai, 1987; Cottle et al., 2013). Wildlife biologists might be expected to know these things; however, a survey of biologists and wildlife workers revealed that many respondents admitted to not using gloves when handling potentially infectious carcasses, and 62% of respondents claimed to have no formal education on zoonotic diseases (Bosch et al., 2013).

Once all parties have been informed of the potential health risks involved in a certain type of fieldwork, training in proper techniques regarding live animal handling, noninvasive sampling, and/or necropsies can further minimize risk (Mills et al., 1995a). Proper understanding of techniques is necessary for the ethical treatment of animals (Sikes et al., 2011; Sikes et al., 2016) as well as the safety of human participants (Mills et al., 1995a). Additionally, thorough use of appropriate disinfectants is one of the most efficient means of preventing the spread of various zoonoses (Mills et al., 1995a). Use of disinfectants on traps has been recommended (Mills et al., 1995a), and despite common concern about small mammals avoiding traps cleaned with chemicals, a recent study (Wilson and Mabry, 2010) found no significant difference in trapping success between treated and untreated traps. Each of these practices (education of risks, medical prophylaxes, familiarity, and the use of appropriate techniques) provides a "layer" of protection to further reduce the risk of exposures.

The proper use of PPE is yet another level of protection. The optimal amount and type of PPE will vary greatly depending upon the type of activity that is planned (Figure 1). For example, a disposable outer layer of clothes (surgical gowns, lab coats, etc.) tucked into your gloves can help reduce the risk of exposure to ectoparasites (Mills et al., 1995a). When entering or cleaning an enclosed area with evidence of small mammal activity or during active handling/necropsy conditions where particles could be aerosolized, the use of respiratory protection is advised (Mills et al., 1995a; Kelt et al., 2010). Either a powered air purifying respirator (PAPR) equipped with a high-efficiency particu-



**Figure 1.** Use of PPE depends upon the situation. The PPE of an Ebola medical responder\* (A) is drastically different than that needed by field researchers conducting necropsies in Colombia (B) or entering caves in Africa (C). 1C photo credit Brian Bird (CDC). \*Indicates acceptable PPE for Ebola medical responders as recommended by WHO.

late arrestance (HEPA) filter, or other HEPA-filtered masks (e.g., N95, if no facial hair is present) in conjunction with a facial splash guard can decrease likelihood of exposure to aerosolized particles and splashes that can transmit a variety of pathogens. Depending on the PAPR type, chemical filters can often be used when working with inhaled anesthetics. However, the purpose of this document is not to provide recommendations or guidelines regarding specific types of PPE, so we defer to institutional requirements and subject matter experts. In addition to the documents cited within this text, a variety of resources are listed in Appendix 1, which offer greater detail regarding selection of appropriate PPE and techniques for minimizing exposure risks.

The use of appropriate PPE has the potential to safe-guard researchers from exposure to infectious agents, but it is not infallible (Fischer et al., 2014). Improper decontamination and doffing techniques can expose the wearer even after active handling has been completed. This suggests that proper training regarding donning and doffing protocols is as important, if not more so, than the presence of PPE itself. A proper discussion and walk-through consisting of multiple doffing techniques can be found at a number of online resources (Appendix 1). It is recommended that doffing procedures be standardized to assure that all participants will use recommended techniques consistently (Fischer et al., 2014).

Given the results of relatively recent surveys of professional organizations and National Park Service employees, it is clear that a large portion of wildlife biologists work consistently without gloves, HEPA masks, or disinfectants (Fulhorst et al., 2007; Kelt et al., 2007; Bosch et al.,

2013), which are some of the least invasive and most effective pieces of PPE available; in addition, many wildlife biologists are not even aware of the need for PPE (Bosch et al., 2013). Reasons cited for the lack of PPE use include items being inconsistently stocked or available, invasive in extreme conditions (such as high heat and humidity), and costly (Bosch et al., 2013). Many of these potential barriers to the use of PPE can be addressed through a number of steps, including increased education regarding potential risks to field biologists, epidemiologists, veterinarians, and other occupations with increased contact with small mammals (e.g., staff at wildlife rehabilitation centers) and allocating a portion of course fees or grant monies toward PPE and proper training. Additionally, professional associations (e.g., The Wildlife Society, Wildlife Disease Association, American Society of Mammalogists, Latin American Mammal Congress) could serve as advocates for cultural change across professions (Bosch et al., 2013). Supervisor (principal investigator [PI]) engagement has been shown to increase compliance with work safety measures (Bosch et al., 2013; Lombardi et al., 2009). The first step toward reducing barriers that impede the use of PPE is to discuss the risks of various types of fieldwork and to consider ways to minimize those risks for all involved. We hope that as new professionals and students prepare to enter the fields of biological and epidemiological research, the topics of biosafety and risk mitigation are fresh on their minds. With knowledge in these areas, researchers in these important fields can continue to increase our understanding of mammalian biodiversity, zoonoses, and epidemiology while responsibly protecting themselves.

**Disclaimer** – The findings and conclusions in this report are those of the authors and do not necessarily represent the views of the Centers for Disease Control and Prevention.

**Acknowledgments** – The authors would like to thank B. R. Amman, C. Hsu, and N. Chea for help acquiring photos of various types of PPE.

#### **Literature Cited**

- Agnandji, S. T., A. Huttner, M. E. Zinser, P. Njuguna, C. Dahlke, J. F. Fernandes, S. Yerly, J. A. Dayer, V. Kraehling, R. Kasonta, and A. A. Adegnika. 2016. Phase 1 trials of rVSV *Ebola* vaccine in Africa and Europe—preliminary report. New England Journal of Medicine 374: 1647–1660. doi:10.1056/NEJMoa1414216
- Albariño, C. G., M. Foltzer, J. S. Towner, L. A. Rowe, S. Campbell, C. M. Jaramillo, B. H. Bird, D. M. Reeder, M. E. Vodzak, P. Rota, and M. G. Metcalfe. 2014. Novel paramyxovirus associated with severe acute febrile disease, South Sudan and Uganda, 2012. Emerging Infectious Diseases 20: 211–216.
- Amman, B. R., C. G. Albariño, B. H. Bird, L. Nyakarahuka, T. K. Sealy, S. Balinandi, A. J. Schuh, S. M. Campbell, U. Ströher, M. E. Jones, and M. E. Vodzack. 2015. A recently discovered pathogenic paramyxovirus, *Sosuga virus*, is present in *Rousettus aegyptiacus* fruit bats at multiple locations in Uganda. Journal of Wildlife Diseases 51: 774–779.
- Baker, W. S., and G. C. Gray. 2009. A review of published reports regarding zoonotic pathogen infection in veterinarians. Journal of the American Veterinary Medical Association 243: 1271–1278.
- Bosch, S. A., K. Musgrave, and D. Wong. 2013. Zoonotic disease risk and prevention practices among biologists and other wildlife workers—results from a national survey, US National Park Service, 2009. Journal of Wildlife Diseases 49: 475–485.
- Bradley, R. D., and M. R. Mauldin. 2016. Molecular data indicate a cryptic species in *Neotoma albigula* (Cricetidae: Neotominae) from northwestern México. Journal of Mammalogy 97: 187–199.
- Brooks, D. R., E. P. Hoberg, W. A. Boeger, S. L. Gardner, K. E. Galbreath, D. Herczeg, H. H. Mejía-Madrid, S. E. Rácz, and A. T. Dursahinhan. 2014. Finding them before they find us: informatics, parasites, and environments in accelerating climate change. Comparative Parasitology 81: 155–164.
- CDC [Centers for Disease Control and Prevention]. 2015. Yellow Book. Centers for Disease Control and Prevention, National Center for Emerging and Zoonotic Infectious Diseases (NCEZID), http://wwwnc.cdc.gov/travel/page/yellowbookhome-2014. Accessed June 26, 2016.
- CDC [Centers for Disease Control and Prevention]. 1993. *Hantavirus* infection—Southwestern United States: interim recommendations for risk reduction. Morbidity and Mortality Weekly Report 42(RR-11): 1–13.
- Ceballos, G. 2014. Mammals of Mexico. Johns Hopkins University Press.

- Ceballos, G., and P. R. Ehrlich. 2006. Global mammal distributions, biodiversity hotspots, and conservation. Proceedings of the National Academy of Science 103: 19374–19379.
- Childs, J. E., T. G. Ksiazek, C. F. Spiropoulou, J. W. Krebs, S. Morzunov, G. O. Maupin, K. L. Gage, P. E. Rollin, J. Sarisky, R. E. Enscore, J. K. Frey, C. J. Peters, and S. T. Nichol. 1994. Serologic and genetic identification of *Peromyscus maniculatus* as the primary rodent reservoir for a new hantavirus in the southwestern United States. Journal of Infectious Diseases 169: 1271–1280.
- Cottle, L. E., E. Gkrania-Klotsas, H. J. Williams, H. E. Brindle, A. J. Carmichael, G. Fry, and N. J. Beeching. 2013. A multinational outbreak of histoplasmosis following a biology field trip in the Ugandan rainforest. Journal of Travel Medicine 20(2): 83–87.
- Davis, R. W., L. A. Fuiman, T. M. Williams, S. O. Collier, W. P. Hagey, S. B. Kanatous, S. Kohin, and M. Horning. 1999. Hunting behavior of a marine mammal beneath the Antarctic fast ice. Science 283: 993–996.
- Delgado, S., B. R. Erickson, R. Agudo, P. J. Blair, E. Vallejo, C. G. Albariño, J. Vargas, J. A. Comer, P. E. Rollin, T. G. Ksiazek, and J. G. Olson. 2008. *Chapare virus*, a newly discovered arenavirus isolated from a fatal hemorrhagic fever case in Bolivia. PLoS Pathogens 4: e1000047.
- Feasey, N., M. Wansbrough-Jones, D. C. Mabey, and A. W. Solomon. 2009. Neglected tropical diseases. British Medical Bulletin 93: 179–200.
- Field, H., P. Young, J. M. Yob, J. Mills, L. Hall, and J. Mackenzie. 2001. The natural history of *Hendra* and *Nipah* viruses. Microbes and Infection 3: 307–314.
- Finnegan, C. J., S. M. Brookes, N. Johnson, J. Smith, K. L. Mansfield, V. L. Keene, L. M. McElhinney, and A. R. Fooks. 2002. Rabies in North America and Europe. Journal of the Royal Society of Medicine 95: 9–13.
- Fischer, W. A., N. A. Hynes, and T. M. Perl. 2014. Protecting health care workers from *Ebola*: personal protective equipment is critical but is not enough. Annals of Internal Medicine 161: 753–754.
- Foy, B. D., K. C. Kobylinski, J. L. Chilson Foy, B. J. Blitvich, A. Travassos da Rosa, A. D. Haddow, R. S. Lanciotti, and R. B. Tesh. 2011. Probable non-vector-borne transmission of *Zika virus*, Colorado, USA. Emerging Infectious Diseases 17: 880–882.
- Fulhorst, C. E., M. D. Bowen, R. A. Salas, N. M. De Manzione, G. Duno, A. Utrera, T. G. Ksiazek, C. J. Peters, S. T. Nichol, E. De Miller, and D. Tovar. 1997. Isolation and characterization of pirital virus, a newly discovered South American arenavirus. American Journal of Tropical Medicine and Hygiene 56: 548–553.
- Fulhorst, C. F., M. L. Milazzo, L. R. Armstrong, J. E. Childs, P. E. Rollin, R. Khabbaz, C. J. Peters, and T. G. Ksiazek. 2007. Hantavirus and arenavirus antibodies in persons with occupational rodent exposure, North America. Emerging Infectious Diseases 13: 532–538.
- Gardner, S. L., J. Salazar-Bravo, and J. A. Cook. 2014. New species of *Ctenomys* Blainville 1826 (Rodentia: Ctenomyidae)

- from the lowlands and central valleys of Bolivia. Special Publication, Museum of Texas Tech University 62: 1–34.
- Gore, M. L., B. A. Knuth, P. D. Curtis, and J. E. Shanahan. 2006. Education programs for reducing American black bear–human conflict: indicators of success? Ursus 17: 75–80.
- Grace, D., F. Mutua, P. Ochungo, R. Kruska, K. Jones, L. Brierley, L. Lapar, M. Said, M. Herrero, P. M. Phuc, and N. B. Thao. 2012. Mapping of poverty and likely zoonoses hotspots. Zoonoses Project 4. Report to the UK Department for International Development. Nairobi, Kenya: International Livestock Research Institute.
- Griffiths, C. M., S. M. Wilson, and J. C. S. Clegg. 1992. Sequence of the nucleocapsid protein gene of *Machupo* virus: close relationship with another South American pathogenic arenavirus, Junin. Archives of Virology 124: 371–377.
- Hemmi, H., T. Kaisho, O. Takeuchi, S. Sato, H. Sanjo, K. Hoshino, T. Horiuchi, H. Tomizawa, K. Takeda, and S. Akira. 2002. Small anti-viral compounds activate immune cells via the TLR7 MyD88–dependent signaling pathway. Nature Immunology 3: 196–200.
- Hoberg, E. P., and D. R. Brooks. 2015. Evolution in action: climate change, biodiversity dynamics, and emerging infectious disease. Philosophical Transactions of the Royal Society B: Biological Sciences 370: 1–7.
- Hutin, Y. J., R. J. Williams, P. Malfait, R. Pebody, V. N. Loparev, S.
  L. Ropp, M. Rodriguez, J. C. Knight, F. K. Tshioko, A. S. Khan,
  M. V. Szczeniowski, and J. J. Esposito. 2001. Outbreak of human *Monkeypox*, Democratic Republic of Congo, 1996 to 1997. Emerging Infectious Diseases 7: 434–438.
- Hutson, C. L., K. N. Lee, J. Abel, D. S. Carroll, J. M. Montgomery, V. A. Olson, Y. Li, W. Davidson, C. Hughes, M. Dillon, and P. Spurlock. 2007. *Monkeypox* zoonotic associations: insights from laboratory evaluation of animals associated with the multi-state US outbreak. American Journal of Tropical Medicine and Hygiene 76: 757–768.
- Irwin, N. R., M. Bayerlova, O. Missa, and N. Martinkova. 2012. Complex patterns of host switching in New World arenaviruses. Molecular Ecology 21: 4137–4150.
- Jones, K. E., N. G. Patel, M. A. Levy, A. Storeygard, D. Balk, J. L. Gittleman, and P. Daszak. 2008. Global trends in emerging infectious diseases. Nature 451: 990–993.
- Kaufman, D. M. 1995. Diversity of New World mammals: universality of the latitudinal gradients of species and bauplans. Journal of Mammalogy 76: 322–334.
- Keckler, M. S., D. S. Carroll, N. F. Gallardo-Romero, R. R. Lash, J. S. Salzer, S. L. Weiss, N. Patel, C. J. Clemmons, S. K. Smith, C. L. Hutson, and K. L. Karem. 2011. Establishment of the black-tailed prairie dog (*Cynomys ludovicianus*) as a novel animal model for comparing smallpox vaccines administered pre-exposure in both high and low-dose *Monkeypox* challenges. Journal of Virology 85: 7683–7698.
- Kelt, D. A., and M. S. Hafner. 2010. Updated guidelines for protection of mammalogists and wildlife researchers from hantavirus pulmonary syndrome (HPS). Journal of Mammalogy 91: 1524–1527.

- Kelt, D. A., D. H. Van Vuren, M. S. Hafner, B. J. Danielson, and M. J. Kelly. 2007. Threat of hantavirus pulmonary syndrome to field biologists working with small mammals. Emerging Infectious Diseases 13: 1285–1287.
- Likos, A. M., S. A. Sammons, V. A. Olson, A. M. Frace, Y. Li, M. Olsen-Rasmussen, W. Davidson, R. Galloway, M. L. Khristova, M. G. Reynolds, H. Zhao, D. S. Carroll, A. Curns, P. Formenty, J. J. Esposito, R. L. Regnery, and I. K. Damon. 2005. A tale of two clades: *Monkeypox* viruses. Journal of General Virology 86: 2661–2672.
- Lombardi, D. A., S. K. Verma, M. J. Brennan, and M. J. Perry. 2009. Factors influencing worker use of personal protective eyewear. Accident Analysis and Prevention 41: 755–762.
- Mann, J. H., O. J. Rollag, H. F. Hull, and J. M. Montes. 1984. Animal bites as an occupational hazard among animal control officers. American Journal of Public Health 74: 255–256.
- Meyers, L. 2007. Contact network epidemiology: bond percolation applied to infectious disease prediction and control. Bulletin of the American Mathematical Society 44: 63–86.
- Mills, J. N., J. E. Childs, T. G. Ksiazek, C. J. Peters, and W. M. Velleca. 1995a. Methods for trapping and sampling small mammals for virologic testing. Atlanta: US Department of Health and Human Services, Public Health Service, CDC.
- Mills, J. N., T. L. Yates, J. E. Childs, R. R. Parmenter, T. G. Ksiazek, P. E. Rollin, and C. J. Peters. 1995b. Guidelines for working with rodents potentially infected with *Hantavirus*. Journal of Mammalogy 76: 716–722.
- Montoya-Ruiz, C., F. J. Diaz, and J. D. Rodas. 2014. Recent evidence of *Hantavirus* circulation in the American tropic. Viruses 6: 1274–1293.
- Moyer, W. M. 2015. The growing battle against blood-sucking ticks. Nature 524: 406–408.
- Murray, K. A., N. Preston, T. Allen, C. Zambrana-Torrelio, P. R. Hosseini, and P. Daszak. 2015. Global biogeography of human infectious diseases. Proceedings of the National Academy of Sciences 112: 12746–12751.
- Musser, G. G., and M. D. Carleton. 2005. Superfamily Muroidea. Pp. 894–1531 in: Wilson, D. E., and D. M. Reeder (eds.). Mammal species of the world. A taxonomic and geographic reference, vol. 2. 3rd ed. Baltimore, MD: Johns Hopkins University Press.
- Myers, N., R. A. Mittermeier, C. G. Mittermeier, G. A. Da Fonseca, and J. Kent. 2000. Biodiversity hotspots for conservation priorities. Nature 403: 853–858.
- Nakazawa, Y., M. R. Mauldin, G. L. Emerson, M. G. Reynolds, R. R. Lash, J. Gao, H. Zhao, Y. Li, J. J. Muyembe, P. M. Kingebeni, and O. Wemakoy. 2015. A phylogeographic investigation of African *Monkeypox*. Viruses 7: 2168–2184.
- Nichol, S. T., C. F. Spiropoulou, S. Morzunov, P. E. Rollin, T. G. Ksiazek, H. Feldmann, A. Sanchez, J. Childs, S. Zaki, and C. J. Peters. 1993. Genetic identification of a *Hantavirus* associated with an outbreak of acute respiratory illness. Science 262: 914–918.
- Nolen, L. D., L. Osadebe, J. Katomba, J. Likofata, D. Mukadi, B. Monroe, J. Doty, J. Malekani, J. Kabamba, P. L. Bomponda,

- and J. I. Lokota. 2015. Introduction of monkeypox into a community and household: risk factors and zoonotic reservoirs in the Democratic Republic of the Congo. American Journal of Tropical Medicine and Hygiene 93: 410–415.
- Olayemi, A., V. Nicolas, J. Hulselmans, A. D. Missoup, E. Fichet-Calvet, D. Amundala, A. Dudu, T. Dierckx, W. Wendelen, H. Leirs, and E. Verheyen. 2012. Taxonomy of the African giant pouched rats (Nesomyidae: *Cricetomys*): molecular and craniometric evidence support an unexpected high species diversity. Zoological Journal of the Linnean Society 165: 700–719.
- Peterson, A. T. 2006. Ecologic niche modeling and spatial patterns of disease transmission. Emerging Infectious Diseases 12: 1822–1826.
- Pitts, R. M., M. R. Mauldin, C. W. Thompson, and J. R. Choate. 2013. Evidence of hantavirus exposure in rodents from north Texas. Western North American Naturalist 73: 386–391.
- Reeder, D. A. M., K. M. Helgen, and D. E. Wilson. 2006. Global trends and biases in new mammalian species discoveries. Occasional Papers, Museum of Texas Tech University 269: 1–35.
- Reynolds, M. G., D. S. Carroll, V. A. Olson, C. Hughes, J. Galley, A. Likos, J. M. Montgomery, R. Suu- Ire, M. O. Kwasi, J. J. Root, Z. Braden, J. Abel, C. Clemmons, R. Regnery, K. Karem, and I. K. Damon. 2010. A silent enzootic of an orthopoxvirus in Ghana, West Africa: evidence for multispecies involvement in the absence of widespread human disease. American Society for Tropical Medicine and Hygiene 82: 746–754.
- Salter, A. J., and B. R. Martin. 2001. The economic benefits of publicly-funded basic research: a critical review. Research Policy 30: 509–532.
- Sikes, R. S., and the Animal Care and Use Committee of the American Society of Mammalogists. 2016. 2016 Guidelines of the American Society of Mammalogists for the use of wild mammals in research and education. Journal of Mammalogy 97: 663–688.
- Sikes, R. S., and W. L. Gannon. 2011. Guidelines of the American Society of Mammalogists for the use of wild mammals in research. Journal of Mammalogy 92: 235–253.

- Sinclair, J. R., D. S. Carroll, J. M. Montgomery, B. Pavlin, K. Mc-Combs, J. N. Mills, J. A. Comer, T. G. Ksiazek, P. E. Rollin, S. T. Nichol, and A. J. Sanchez. 2007. Two cases of *Hantavirus* pulmonary syndrome in Randolph County, West Virginia: a coincidence of time and place? American Journal of Tropical Medicine and Hygiene 76: 438–442.
- Torres-Pérez F., L. Wilson, S. K. Collinge, H. Harmon, C. Ray, R. A. Medina, and B. Hjelle. 2010. Sin Nombre virus infection in field workers, Colorado, USA. Emerging Infectious Diseases 16: 308–310.
- Tsai, T. F. 1987. Hemorrhagic fever with renal syndrome: mode of transmission to humans. Laboratory Animal Science 37:428–430.
- van der Straeten, E., J. Kerbis Peterhans, K. Howell, and N. Oguge. 2008. *Cricetomys gambianus*. The IUCN Red List of Threatened Species 2008: e.T5522A11272093. doi: org/10.2305/IUCN.UK.2008.RLTS.T5522A11272093.en. Downloaded April 1, 2016.
- WHO [World Health Organization]. 2005. WHO Expert Consultation on Rabies, Geneva. WHO Technical Report Series No. 931.
- Willig, M. R., D. M. Kaufman, and R. D. Stevens. 2003. Latitudinal gradients of biodiversity: pattern, process, scale, and synthesis. Annual Review of Ecology, Evolution, and Systematics 34: 273–309.
- Wilson, J. A., and K. E. Mabry. 2010. Trapping in a cautious world: the effect of disinfectants on trap success. Western North American Naturalist 70: 467–473.
- Wong, D., M. A. Wild, M. A. Walburger, C. L. Higgins, M. Callahan, L. A. Czarnecki, E. W. Lawaczeck, E. Craig, J. Levy, G. Patterson, R. Suneshine, and P. Adem. 2009. Primary pneumonic plague contracted from a mountain lion carcass. Clinical Infectious Diseases 49: e33–38.
- Young, T. P. 1994. Natural die-offs of large mammals: implications for conservation. Conservation Biology 8: 410–418.
- Zeitz, P. S., J. C. Butler, J. E. Cheek, M. C. Samuel, J. E. Childs, L. A. Shands, R. E. Turner, R. E. Voorhees, J. Sarisky, P. E. Rollin, and T. G. Ksiazek. 1995. A case-control study of *Hantavirus* pulmonary syndrome during an outbreak in the southwestern United States. Journal of Infectious Diseases 171: 864–87

## Appendix I

A list of resources (in addition to those cited within the text) for more detailed information regarding biosafety in the field. These references include information from specific pathogens endemic to certain countries and prevention/treatment measures, to donning/doffing procedures, and selection of appropriate PPE.

- Mills, J. N., Carroll, D. S., Revelez, M. A., Amman, B. R., Gage, K. L., Henry, S., & Regnery, R. L. (2007).
   Minimizing infectious disease risks in the field. Wildlife Professional, 1(4), 30.
- CDC Traveler's Health page: contains information regarding endemic diseases, ongoing outbreaks, special prophylaxis recommendations for countries around the world: <a href="http://wwwnc.cdc.gov/travel/">http://wwwnc.cdc.gov/travel/</a>
- CDC Yellowbook: updated annually can be purchased as a hard copy or electronic version. For more
  information visit website <a href="http://wwwnc.cdc.gov/travel/page/yellowbook-home-2014">http://wwwnc.cdc.gov/travel/page/yellowbook-home-2014</a>
- WHO country information (<a href="http://www.who.int/countries/en/">http://www.who.int/countries/en/</a>)
- Armed Forces Pest Management Board Technical Guide No. 41: Protection from rodent-borne diseases
  with emphasis on occupational exposure to hantavirus: contains information regarding a variety
  of pathogens, their reservoirs, geographic distributions, and routes of transmission, as well as risk
  mitigation for hantaviruses <a href="http://www.afpmb.org/sites/default/files/pubs/techguides/tg41.pdf">http://www.afpmb.org/sites/default/files/pubs/techguides/tg41.pdf</a>
- CDC veterinary safety and health website: includes information on various zoonoses, links to other sites
  and documents for information on PPE, biological waste disposal:
   <a href="http://www.cdc.gov/niosh/topics/veterinary/biological.html#waste">http://www.cdc.gov/niosh/topics/veterinary/biological.html#waste</a>
- The online version of the Biosafety in Microbiological and Biomedical Laboratories (BMBL 5<sup>th</sup> edition) includes information regarding various agents for use in sterilization and disinfection of spaces and surfaces, as well as use of appropriate biosafety in laboratory conditions. <a href="http://www.cdc.gov/biosafety/publications/bmbl5/">http://www.cdc.gov/biosafety/publications/bmbl5/</a>
- An example of an Institutional Animal Care and Use Committee Standard Operating Procedure from the University of Colorado, Boulder website: <a href="http://www.colorado.edu/vcr/sites/default/files/attached-files/SOP%2015%20PPE%20Policy%20">http://www.colorado.edu/vcr/sites/default/files/attached-files/SOP%2015%20PPE%20Policy%20</a> 08072013\_0.pdf
- The National Park Service's page for Vectorborne & Zoonotic Infectious Agents includes links for information on various pathogens, their vectors, potential hosts, and safe practices to avoid zoonotic disease from wildlife: <a href="https://www.nps.gov/public\_health/di/vb\_ia.htm">https://www.nps.gov/public\_health/di/vb\_ia.htm</a>.
- World Health Organization's Rapid advice guidelines: Personal protective equipment in the context of filovirus disease outbreak response.
   <a href="http://apps.who.int/iris/bitstream/10665/137410/1/WHO\_EVD\_Guidance\_PPE\_14.1\_eng.pdf?ua=1">http://apps.who.int/iris/bitstream/10665/137410/1/WHO\_EVD\_Guidance\_PPE\_14.1\_eng.pdf?ua=1</a>